Using plant growth promoting rhizobacteria for improving tomato growth under saline stress

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Abstract

Salinity is a crucial constraint that slow downs agriculture production in many areas in Egypt. Inoculation with plant growth promoting microorganisms may enhance plant growth under salt stress conditions. The objective is to evaluate the inoculation efficieny of biostimulant strains (*Pseudomonas fluorescence D23*, *Bacillus punilus D139* and *Azospirillum lipoferum D178*), humic acid and organic manure (compost) on growth and yield of tomato (*Solanum lycopersicum* L.). This experiment was conducted in greenhouse conditions at the Experimental Farm Station of Faculty of Agricuture Moshtohor during 2011. The highest significant increase of dehydrogenase, nitrogenase and phosphatase was observed in tomato inoculated with biostimulant combined with humic acid + compost at one and half dose. The highest records of macronutrients uptake by tomato shoots were observed when tomato amended with biostimulant combined with compost at different doses + humic acid. Application of humic acid combined with compost significantly decreased the proline content in tomato, whereas, the reverse was observed in nitrate reductase. Dual treatment of tomato with biostimulant and compost gave higher records of tomato growth characteristics and yield.

Key words: Compost, nitrate reductase, plant growth-promoting rhizobacteria, PGPR, praline, saline stress, tomato.

Introduction

Vegetables are important protective food and highly beneficial for the maintenance of health and prevention of diseases. Tomato is a major vegetable crop that has achieved tremendous popularity over the last century. Salinity is one of the most critical constraints which hampers agriculture production in many areas around the world, including Egypt. About 9.5 billion ha of the world's soil are saline, except for large areas of secondary salinized soil in cultivated land (Aşik *et al.*, 2009). Out of 14.12 million ha of arable area of Egypt, 4.2 millions are salt affected. Most of these lands are annually lost for cultivation due to salinity. It is hypothesized that the use of plant growth promoting microorganisms as inoculants can enhance plant growth under salt stress conditions (Nadeem *et al.*, 2006). Plant growth promoting rhizobacteria (PGPR) are free living soil-borne bacteria or a symbiotic one which colonize the rhizosphere. These bacteria enhance plant growth either by direct or indirect mechanisms (Pallai, 2005). The aim of this research is to study the inoculation efficiency of tomato with salttolerant PGPR combined with compost and/or humic acid on tomato growth performance, productivity and yield quality.

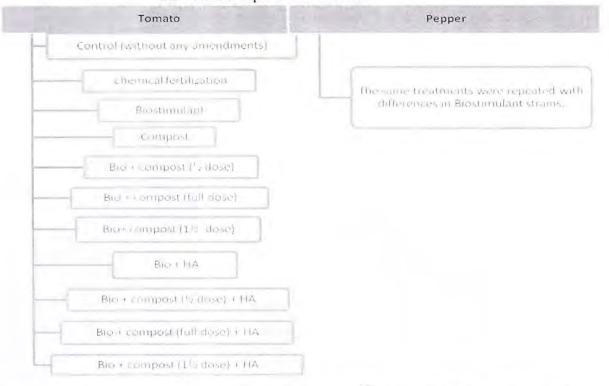
Table 1. T	The experimental	soil texture and	chemical analyses.
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Parameters		Unit	Val	ues
	Soil texture		Soil before adjustment	Soil after adjustmen
Sand		(%)	45.01	50
Silt		(%)	16.32	33
Clay		(%)	38.67	17
Textural class		(%)	Clay	Clayey silt
	Chemical analy	/sis		
EC		ds m ⁻¹	24.34	13.92
pH			8.30	8.61
Organic matter		(%)	0.62	1.12
Total nitrogen		(%)	0.010	0.114
	$Na^+ + K^+$		1.34	8.27
Soluble cations	Ca ₂	meq L ⁻¹	14.22	35.0
	Mg ²		27.12	38.0
	$CO_3^{=}$		Zero	Zero
Coluble enione	HCO ₃	man I -l	9.93	28.00
Soluble anions	CI	meq L ⁻¹	186	81.00
	SO_4^2		47.75	30.20

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Experimental design

Treatments were distributed in a randomized complete block design with three replicates. Flow chart of experiment treatments



adhesive agent. The same prepared PGPR inocula were added to grown plants three times throughout the growing season at a rate of 300 mlpot⁻¹. The recommended dose (full dose) of compost was 8-10 ton fed⁻¹ for vegetable crops. Whereas, humic acid was added to soil at rate of 3-4 kg fed⁻¹. A half dose of inorganic nitrogen fertilizer (50 kg N fed⁻¹) as ammonium sulphate was supplemented for treatments of biostimulant and biostimulant + humic acid. Also, a full dose of inorganic phosphorus fertilizer (25 kg P_2O_3 fed⁻¹) as calcium superphosphate and potassium fertilizer (40 kg K₂O fed⁻¹) as potassium sulphate were supplemented for all treatments.

Materials and methods

Soil and soil mixtures

Soil of the experiment was obtained from Port Said Governorate, Sahl El- Hussinia. Experimental soil was subjected to adjustment with soil gypsum and sand, analyses were carried out according to the method described by Page *et al.* (1982).

Determination

Dehydrogenase activity was assayed in soil according to Glathe' and Thalmann (1970). Phosphatase activity was estimated according to Drobnikova (1961). Nitrogenase activity was Mechanical (particle size distribution) and chemical analyses are presented in Table 1.

Preparation of biostimulant inocula

The biostimulant inocula for tomato (*Pseudomonas fluorescence D23, Bacillus pumilus D139 and Azospirillum lipoferum D178*) were prepared in specific broth media. Cell suspension of *A. lipoferum* contains about (1 x 10^6 cfu ml⁻¹) 7 days-old, *B. pumilus* (90 x 10^6 cfu ml⁻¹) 2 days-old and *Ps. fluorescence* (20 x 10^6 cfu ml⁻¹) 5 days-old.

Cultivation process

Prior to transplanting, tomato seedlings were soaked by dipping the root system in a mixture of PGPR inocula (cell suspension of biostimulant) for 60 minutes; sucrose solution (40 %) was used as an measured by using the acetylene reduction technique given by Diloworth (1970).

Growth and yield traits

Leaves number, flowers number, dry weights of the plants were determined at flowering stage (60 days), plant height was determined after 120 days of transplanting. Number of fruits plant⁻¹, fruits yield and weight of fruit plant⁻¹ were estimated.

Macro-element content

Total nitrogen, phosphorus and potassium contents were determined according to the methods described by (A.O.A.C, 1970); A.P.H.A. (1992) and Dewis and Freitas (1970), respectively.

Proline and nitrate reductase

Proline was determined according to the method of Bates et al. (1973). Nitrate reductase was determined using the method of Abdel-Samad et al. (2004).

Statistical analysis was carried out according to Snedecor and Cochran (1989). The differences between the means value of various treatments were compared by Duncan's multiple range test (Duncan's, 1955).

Results and discussion

Effect of tomato inoculation with biostimulants in presence of compost and/or humic acid on the activity of

Dehydrogenase (DHA) is shown in Fig.1. The rhizosphere of tomato cultivated in salt-affected soil with no amendments (control) gave lower DHA values, this may be due to the high salt concentration which decreases the microbial activities. The DHA in various treatments were significantly higher at flowering stage (60 days) than vegetative one. This increase of DHA could be attributed to the beneficial effect of root exudates which increase during flowering stage. Higher activity of DHA at flowering stage is likely to be due to the higher multiplication rate of different soil microorganisms.

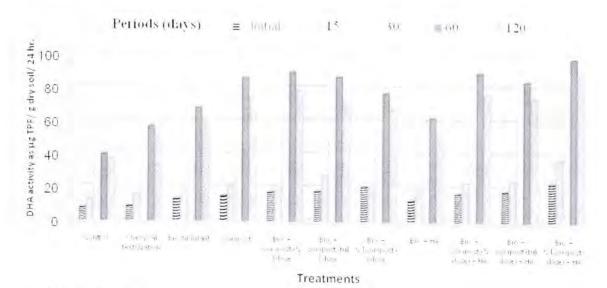


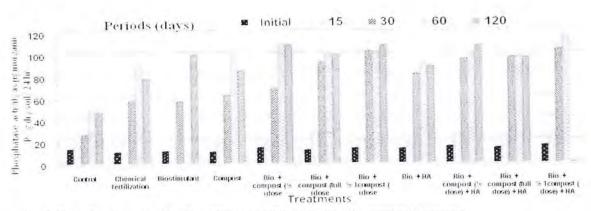
Fig 1. Periodical changes in dehydrogenase activity in soil cultivated with tomato.

Data in Fig. 1 also revealed that the highest significant increase of DHA was observed in case of tomato inoculated with biostimulant combined with humic acid and compost at one and half dose. This result is likely to be due not only to the promotion effect of biostimulant on microbial proliferation but also to the beneficial effect of compost and humic acid. Liu et al. (1992) reported that the addition of humic acid to soil enhanced DHA at vegetative and flowering stages. This result could be attributed to the synergistic effect between biostimulant and natural microbial flora occurred in compost which increased the microbial respiration rate. Obtained data showed that relative lower records of DHA were observed in soil amended with chemical fertilization than soil amended with humic acid and/or compost. This result was observed with most experimental periods and was in accordance with Marinara et al. (2000) who reported that higher

DHA values were observed in soil amended with compost compared to soil fertilized with chemical fertilizers.

Phosphatase activity

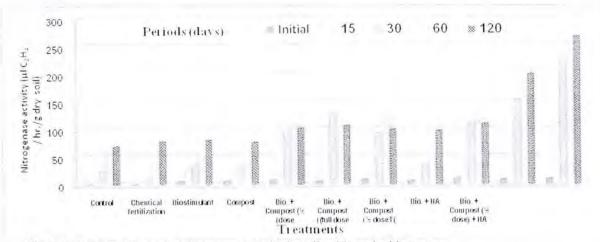
Data illustrated in Fig. 2 showed that inoculation of tomato with *Pseudomonas fluorescence D23*, *Bacillus pumilus D139* and *Azospirillum lipoferum D178* resulted in a significant increase of phosphatase activity compared with either the compost or the chemical fertilization. Obtained results revealed that no significant difference was observed between phosphatase activity in soil treated with biostimulant only and soil treated with chemical fertilizers at 15 and 30 days. Soil inoculated with biostimulant combined with humic aid + compost at one and half dose gave the highest significant values of phosphatase activity.

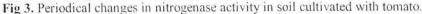


Using plant growth promoting rhizobacteria....

Fig 2. Periodical changes in phosphatase activity in soil cultivated with tomato.

This result may be attributed not only to the effect of inoculation on microbe's number in rhizosphere but also to the beneficial effect of compost on indigenous and introduced biostimulant strains for proliferation and their activities. Balakrishnan *et al.* (2007) found that the application of compost in combination with phosphate solubilizing bacteria significantly increased the soil enzyme activities such as phosphatase. Also, Bama *et al.* (2008) applied humic acid at 20 or 10 kg ha⁻¹ with foliar spray and recorded an increase of enzymatic activities such as catalase, dehydrogenase and phosphatase. Data also showed that phosphatase activity was significantly higher at flowering stage than vegetative one. In case of nitrogenase activity (N2-ase) (Fig. 3), it was affected by the investigated treatments. Tomato amended with chemical fertilizers gave the lowest values of N₂-ase activity compared to other treatments. This result is in harmony with that obtained by Anne-Sophie *et al.* (2002) who found that the addition of chemical fertilizers such as ammonium nitrate decreased the nitrogenase activity.





amended with compost at one and half dose and humic acid in combination with biostimulant. This result may be due to the enhancement of humic acid to the native and introduced microorganisms and also increased the synergistic effect of inocula addition with other microorganisms. These results are in harmony with Meunchang *et al.* (2006) who mentioned that compost promotes plant growth when amended with N₂ - fixing bacteria. Also, data revealed that soil without any amendments gave significant higher nitrogenase activity than soil treated with NPK fertilizers. It may be due to the activity of native microorganisms. Higher records of N₂-ase activity were observed in soil treated with compost than biostimulant each one singularly. Enhancement of biological activities caused by compost might be due to containing native microorganisms. The highest significant values of N₂-ase activity were observed in soil

Treatment	Leaf number plant ⁻¹	Flower number plant ⁻¹	Shoot length	Root dry weight	Shoot dry weight
	[10	Cm		-g
Control ^a	12 ^g	10 ^f	42'	8.9 ^a	15.7ª
Chemical fertilization	30 ^b	24^{ab}	86 ^a	7.8 ^{ab}	14.5 ^b
Biostimulant (Bio) ^b	15 ^{er}	13er	44 ^{et}	5.4 ^c	10.9 ^d
Compost	19^{d}	15 ^{de}	46 ^{ef}	7.4 ^{ab}	8.00 ^c
Bio .+ compost (1/2 dose)	14^{lg}	19 ^{cit}	51 ^{de}	8.4 ^a	6.50 ^f
Bio .+ compost (full dose)	17de	19 ^{cd}	54 ^{cd}	6.7 ^{bc}	10.8^{d}
Bio. + compost (11/2 dose)	22°	19 ^{cat}	50 ^{de}	6.8 ^{bc}	12.9 ^c
Bio .+ HA ^c	15 ^{el}	14 ^{er}	54 ^{cd}	7.2 ^{ab}	8.20 ^e
Bio. + compost ($\frac{1}{2}$ dose) + HA	30 ^b	21 ^{be}	58 ^c	7.0 ^{ab}	14.4 ^b
Bio.+ compost (full dose) + HA	30 th	26°	57 ^{ed}	6.5 ^{bc}	14.0 ^{bc}
Bio. + compost (1½ dose) + HA	40^{a}	26°	70 ^b	7.8 ^{ab}	15.3ª

Table 2. Growth characteristics of tomato planted in salt-affected soil in response to different soil amendements.

"Control:=without any soil amendments.

^bBiostimulant strains = Ps. fluorescence D23, B. pumilus D135 and A. lipoferum D178, ^cHA= Humic acid

Growth characteristics

Data in Table 2 clearly indicated that the lowest records of tomato growth characteristics i.e. leaves number, flowers number and shoot length were observed in plants cultivated in salt-affected soil without any amendments (control). Dual treatment of tomato with biostimulant strains and compost gave higher records of growth characteristics than plants cultivated in soil treated with either biostimulant or compost only. This might be due to the synergistic effect of compost and biostimulant (Table 2).

Significant increase in tomato growth characteristics was observed in soil treated with chemical fertilizers than soil treated with biostimulant only. Respecting the interaction effect between the biostimulant and compost amendment, results revealed that the combination of biostimulant with compost at one and half dose gave high records of tomato shoot length. Similar results were observed by Meunchang et al. (2006) who mentioned that the compost promote plant growth when it was amended with N2-fixing bacteria because the N2-fixing bacteria colonize roots when compost was used and enhance shoot and root growth. Also, Ranganathan et al. (1995) demonstrated that inoculation of tomato seedlings with Azospirillum spp. increased growth, flowering and dry matter of plants. It is worthy to mention that leaf number was not affected by the amount of compost, therefore tomato grown in soil amended with compost at half, full and one and half dose gave similar records. Moreover, when soil amended with compost at different doses in presence of humic acid and biostimulant tomato gave higher values of most determined characteristics than other treatments.

Whereas, at 120 days soil amended with biostimulant gave higher values of N₂-ase activity than compost only, this result explained the importance of the poost inocula which added to the experimental soil. Obtained data in Fig. 3 showed that soil treated with compost in combination with biostimulant gave significant higher values of N₂-ase activity in tomato rhizosphere than soil solely treated with each only.

Nitrogen, phosphorus and potassium uptake

Results in Table 3 showed that the lowest values of macronutrients uptake were observed in saltaffected soil without any amendments (control). This may be attributed to the negative effect of salinity on macronutrients absorption. These results are in harmony with Lopez and Satti (1996) who proved that salinity can reduce N2 accumulation in plants, P concentrations and the uptake of K in plants due to the inhibitive effect of Na on such process. Obtained data revealed that N, P and K uptake were significantly increased in plants grown in soil treated with chemical fertilization than soil amended with compost or biostimulant strains, each one. Moreover, N, P and K uptake by tomato shoots were higher in case of dual application with biostimulant and compost than those recorded in either biostimulant or compost solely with each one. This may be due to the beneficial effect of dual application on macronutrients availability and uptake. The highest records of macronutrients uptake by tomato shoots were observed when soil was amended with biostimulant combined with compost at different doses in presence of humic acid. This might be due to the positive effect of compost on chemical properties of salt-affected soil which might cause release of macronutrients or availability of nutrients. Similar results were

observed by Alvarez et al. (1995) who reported that the addition of compost to soil cultivated with tomato may affect the release of nutrients to plants directly through the inherent nutrients or indirectly by their effect on the cation-exchange capacity.

Table 3. Uptake of N	, P and K by tomato o	cultivated in salt-affected soil	l in response to soil amendements.
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Treatment	N	Р	K
		mg plant ¹	
Control ^a	66.58 ^c	8.25°	31.82 ^b
Chemical fertilization	92.84 ^b	13.3ª	43.88ª
Biostimulant (Bio) ^b	80.16 ^{bc}	9.25 ^b	40.62°
Compost	84.91 ^{bc}	9.80 ^b	38.90 ^{ab}
Bio. + compost ($\frac{1}{2}$ dose)	89.67 ^{bc}	11.1 ^{ab}	35.26 ^{ab}
Bio. + compost (full dose)	90.89 ^b	11.2 ^{ab}	36.68 ^{ab}
Bio. + compost $(1\frac{1}{2} \text{ dose})$	98.98 ^b	11.4^{ab}	38.78 ^{ab}
Bio. $+ HA^{c}$	89.16 ^{bc}	11.0 ^{ab}	36.00 ^{ab}
Bio. + compost ($\frac{1}{2}$ dose) + HA	108.9 ^{ab}	13.4^{a}	33.64 ^b
Bio. + compost (full dose) + HA	118.4 ^{ab}	13.6°	43.04ª
Bio. + compost (11/2 dose) + HA	123.9 ^a	14.6 ^a	44.80 ^a

"Control:=without any soil amendments.

^bBiostimulant strains = Ps. fluorescence D23, B. pumilus D135 and A. lipoferum D178, ^cHA= Humic acid

In addition, soil treated with humic acid gave higher values of N, P and K uptake by tomato shoots compared with control. This is due to the beneficial effect of humic acid application on the decrease of soil salinity. This result is in agreement with Masciandaro *et al.* (2002) who found that humic substances may enhance the nutrients uptake and reduce the uptake of some toxic elements. Therefore, it could be said that the application of humic substances could improve plant growth under salinity conditions.

Proline accumulation and nitrate reductase activity

Data recorded in Table 4 clearly indicated that saltaffected soil without any amendments (control) gave the highest amounts of proline in tomato plants. It may be due to the response of plants to high concentration of salts. Tomato inoculated with biostimulant combined with compost at one and half dose in presence of humic acid gave the lowest amounts of accumulated proline. This result could be attributed to the beneficial effect of natural microbial flora occurred in compost or biostimulant which have been demonstrated to induce plant tolerance to salinity. This result is in agreement with Mohamed et al. (2007) who reported that proline content significantly increased with an increase of NaCl concentration. Also, Martinez et al. (1996) found a positive relationship between proline accumulation and NaCl tolerance. In addition, the application of humic acid combined with compost significantly decreased the accumulated proline amounts in tomato plants rather than the absence of humic acid. It may be likely due to the role of humic acid in decreasing the effect of salinity on plants. Humic acid could be used as a growth regulator to regulate hormone level, improve plant growth and enhance stress tolerance (Serenella et al., 2002). Results clearly indicated that except for the control, chemical fertilization treatment gave the lowest values of nitrate reductase (NR-ase).

 Table 4. Proline accumulation and nitrate reductase activity in tomato leaves in salt-affected soil in response to soil amendements.

Treatment	Proline accumulation	Nitrate reductase activity
	mg g ⁻¹	μ mol NO ₂ g ⁻¹ hr ⁻¹
Control ^a	0.84^{a}	194.7 ^a
Chemical fertilization	0.82^{a}	165.0°
Biostimulant (Bio) ^b	0.72 ^b	177.2 ^b
Compost	0.45 ^c	141.3 ^d
Bio. + compost (1/2 dose)	0.28°	76.10 ^g
Bio. + compost (full dose)	0.38^{d}	99.00 [°]
Bio. + compost (11/2 dose)	0.45°	100.5 ^r
$Bio. + HA^c$	0.79^{3}	170.6 ^c
Bio. + compost (1/2 dose) + HA	0.49^{c}	52.70'
Bio. + compost (full dose) + HA	0.51 ^c	58.90 ^h
Bio. + compost (1½ dose) + HA	0.45 ^c	123.5°

^bBiostimulant strains = Ps. fluorescence D23, B. pumilus D135 and A. lipoferum D178, ^c HA= Humic acid

Microbial and nathological bio-techniques.

^aControl:=without any soil amendments.

inoculations with biostimulant in Tomato combination with compost at different doses significantly increased the nitrate reductase activity compared to either biostimulant or compost each one individually. Also, nitrate reductase increased with the increasing of compost amounts since the addition of compost at one and half dose gave higher records than either half or full dose. The highest values of NR-ase were observed in case of soil amended with compost at one and half dose in presence of humic acid and biostimulant. This result may be due to the beneficial effect of the native microorganisms occurred in compost and their synergistic effect with biostimulant. These results are in agreement with Zhang et al. (2008) who stated that the positive effects of humic acid on plant growth could be mainly due to hormone-like activities of the humic acid through their involvement in oxidative phosphorylation, protein synthesis, antioxidant and various enzymatic reactions.

Yield and yield components

Data in Table 5 revealed that the lowest number of fruits/plant was observed in tomato grown in salt-affected

soil without any amendments (control), while the treatments that contain biostimulant, compost and/or humic acid gave higher number of fruits plant¹. Similar results were observed by Ullah *et al.* (1994) who reported that tomato fruit production was negatively affected by high salt concentrations. Results

also showed that the number of fruits plant¹ was similar in case of tomato inoculation with biostimulant combined with compost at different doses in presence of humic acid. This trend of results is in agreement with Zandonadi et al. (2007) who reported that humic acid increased growth and yield of various crops including vegetables. Chemical fertilization of tomato gave significant higher number of fruits than tomato inoculation with either biostimulant or compost. The highest weight of tomato fruits was observed when tomato inoculated with biostimulant in combination with compost at one and half dose in presence of humic acid. Obtained data in Table 5 also revealed that tomato yield was significantly increased with the increasing of compost dose, it may be due to the synergistic effect of natural microbial flora occurred in compost on the introduced inocula (biostimulant). Chemical fertilization of tomato gave higher yield of fruits plant¹ than tomato inoculated with either biostimulant or compost. The highest significant yield of tomato fruits was observed when tomato was inoculated with biostimulant in combination with compost at one and half dose in presence of humic acid. This result could be due to the beneficial effects of humic acid and compost. These results are in harmony with Zhang et al. (2008) who reported that the positive effects of humic acid on productivity, which seem to be concentration-related. could mainly be due to hormone like activities of the humic acid through their involvement in cell respiration. photosynthesis, oxidative phosphorylation, protein synthesis, antioxidant and various enzymatic reactions.

Table 5. Yield and yield components of tomato grown in salt-affected soil in response to s	oil
amendements.	

Treatments	Number of fruits plant ¹	Weight of one fruit	Fruits yield plant
	no	g	kg
Control ^a	9 ^b	45.7 ^d	0.411 ^r
Chemical fertilization	1 S ^{ab}	50.7 ^{bcd}	0.761 ^{cd}
Biostimulant (Bio) ^b	12^{ab}	53.8 ^{abc}	0.646 ^e
Compost	14^{ab}	51.5 ^{bed}	0.721^{de}
Bio. + compost (1/2 dose)	14 ^{ab}	54.2 ^{abc}	0.759 ^{ed}
Bio. + compost (full dose)	15 ^{ab}	57.8 ^a	0.867^{ab}
Bio. + Compost (11/2 dose)	15 ^{ab}	55.5 ^{ab}	0.833 ^{bc}
Bio. $+ HA^{c}$	14^{ab}	49.1 ^{cd}	0.687 ^{de}
Bio. + compost (1/2 dose) + HA	16°	59.1 ^a	0.946^{a}
Bio .+ compost (full dose) + HA	16^{a}	58.3 ^a	0.934ª
Bio. + compost $(1\frac{1}{2} \text{ dose})$ + HA	16^{a}	59.7ª	0.955 ^a

^aControl:=without any soil amendments.

^bBiostimulant strains = Ps. fluorescence D23, B. pumilus D135 and A. lipoferum D178, ^cHA= Humic acid

Also, Ulukan (2008) reported that the improving soil conditions and establishing equilibrium among plant nutrients are also important for soil productivity and plant production. Moreover, humic substances and organics improve the soil characteristics and increase the yield of vegetable crops.

Microbial and pathological bio-techniques,

Conclusion

This study recommended that the use of salt tolerant PGPR as biostimulant for tomato grown in saltaffected soils can improve growth performance and productivity under saline stress. Results also indicated that tomato inoculation with PGPR in combination with compost and humic acid enhanced plant resistance to stress through the reduction of proline accumulation and increase of nitrate reductase.

References

- A.P.H.A. American Public Health Association. 1992. Standard methods for the examination of water and waste water. Washington, DC, U.S.A.
- A.O.A.C., Association of Official Agriculture Chemists 1980. Official methods of analysis 10th Ed. Washington, D.C., USA, P: 832.
- Abdel-Samad, Hamdia; M.A.K. Shaddad and Doaa, M. Mohamed. 2004. Mechanisms of salt tolerance and interactive effects of *Azospirillum brasilense* inoculation on maize cultivars grown under salt stress conditions. Plant Growth Reg., 44:165-174.
- Alvarez, M.A.B.; S. Gagne and H. Antoun. 1995. Effect of compost on rhizosphere microflora of the tomato and on the incidence of plant growthpromoting rhizobacteria. Appl. Environ. Micro., 61(1):194-199.
- Anne-Sophie, V.; S. Christophe; G. M. Nathalie and N. Bertrand. 2002. Quantitative effects of soil nitrate, growth potential and phenology on symbiotic nitrogen fixation of pea (*Pisum* sativum L.).Plant and Soil, 243: 31–42.
- Aşik, B.B.; M.A. Turan; H. Çelik and A.V. Katkat. 2009. Effects of humic substances on plant growth and mineral nutrients uptake of wheat (*Triticum durum* cv. salihli) under conditions of salinity. Asian J. Crop Sci., 1(2):87-95.
- Balakrishnan, V.; K. Venkatesan and K.C. Ravindran 2007. The influence of halophytic compost, farmyard manureand phosphobacteria on soil microflora and enzyme activities Plant Soil Environ., 53(4): 186–192.
- Bama, S.; K. Somasundaram; S.S. Porpavai; K.G. Selvakumari and T.T. Jayaraj. 2008. Maintenance of soil quality parameters through humic acid application in an alfisal and inceptisol. Aust. J. Basic Appl. Sci., 2:521-526.
- Bates, L.S.; R.P. Waldren and I.D. Teare. 1973. Rapid determination of free proline for water stress studies. Plant Soil, 39(1): 205-207.
- Dewis, G. and F. Freitas. 1970. Physical and chemical methods of soil and water analysis. FAO, Bull., No (10).
- Diloworth, M.J. 1970. The acetylene reduction method for measuring biological nitrogen fixation. Rhizobium News Letters, 15(7): p 155.

- Drobnikova, V. 1961. Factors influencing the determination of phosphatase in soil. Folia. Microbiol., 6: 260.
- Duncan's, D. B. 1955. Multiple range and multiple F. test. Biometrics, 11: 11-24.
- Glathe', H. and A. Thalmann .1970. Uber die microbiello activitat and iher Beziehungen zu Fruchtbrkeitsmerkmalen einiger Acherboden unter besonderer Berucksichtigung der dehydrogenase akativitat (TCC. Redukation). Zbl. Bakt. Abt. II, 124: 1-23.
- Liu, S.T.; L.Y. Lee; C.Y. Tai; C.H. Hug; Y.S. Chang; J.H. Wolfram; R. Rogers and A.H. Goldstein. 1992. Cloning of an *Erwinia herbicola* gene necessary for gluconic acid production and enhanced mineral phosphate solubilization in *E. coli* HB101. J. Bacteriol., 174:5814-5819.
- Lopez, M.V. and S.M.E. Satti.1996. Calcium and potassium- enhanced growth and yield of tomato under sodium-chloride stress. Plant Sci., 114:19-27.
- Marinara, S.; G. Masciandro; B. Ceccanti and S. Grego 2000. Influence of organic and mineral fertilizers on soil biological and physical properties. Bioresour. Technol., 72:9-17.
- Martinez, C.A.; M. Maestri and E.G. Lani. 1996. In vitro salt tolerance and proline accumulation in Andean potato (Solanum spp.) differing in frost resistance. Plant Sci., 116:177-184.
- Masciandaro, G.: B. Ceccanti; V. Ronchi; S. Benedicto and L. Howard. 2002. Humic substances to reduce salt effect on plant germination and growth. Commun. Soil Sci. Plant Annals, 33:365-378.
- Meunchang, S.; S. Panichsakpatana and R.W. Weaver 2006. Tomato growth in soil amended with sugar mill by-products compost. Plant Soil, 280:171-176.
- Mohamed, A.N.; M.H. Rahman; A.A. Alsadon and R. Islam. 2007. Accumulation of proline in NaCl-treated callus of six tomato (*Lycopersicon esculentum* Mill.) cultivars. Plant Tissue Cult. Biotech., 17(2):217-220.
- Nadeem, S.M; I. Hussain; M. Naveed; H.N. Asghar; Z.A. Zahir and M. Arshad. 2006. Performance of plant growth promoting rhizobacteria containing ACC-deaminase activity for improving growth of maize under salt-stressed conditions. Pak. J. Agric. Sci., 43(3-4):114-121.
- Page, A. L.; R. Miller and H. Keeney. 1982. Methods of soil analysis. Part 2, 2nd Ed., Am. Soc. Agronomy, Inc. Mad. Wisconsin, USA.
- Pallai, R. 2005. Effect of plant growth-promoting rhizobacteria on canola (*Brassica napus* L.) and lentil (*Lens culinaris* medik.) plants. M.Sc. Thesis, Sasskatoon, Canada.
- Ranganathan, D.S.; R. Perumal and R. Perumal. 1995. Effect of micronutrients without organics and bio-fertilizers on growth and development of

Using plant growth promoting rhizobacteria....

tomato in inceptisol and alfisol. South-Indian Hort., 43(3-4): 89-92.

- Serenella, N.; D. Pizzeghelloa; A. Muscolob and A. Vianello. 2002. Physiological effects of humic substances on higher plants. Soil Biol. Biochem., 34:1527-1536.
- Snedecor, G.W. and W.G. Cochran. 1989. Statistical methods. 8th Ed. Iowa State Univ. Press. Ames Iowa, USA.
- Ullah, S. M.; M.H. Gerzabek and G. Soja (1994). Effect of sea water and soil salinity on ion uptake, yield and quality of tomato (fruit). Die Bodenkulture, 46:227-237.
- Ulukan, H. 2008. Humic acid application into field crops cultivation. Kahraman Maras Sutcu Imam Univ. J. Sci. Eng., 11(2):119-128.
- Zandonadi, D.B.; L.P. Canellas and A.R. Facanha. 2007. Indole acetic and humic acid induce lateral root development through a concerted plasma lemma and tonoplast H[°] pumps activation. Planta, 225:1583-1595.
- Zhang, X.; E.H. Ervin and R.E. Schmidt. 2008. Plant growth regulators can enhance the recovery of Kentucky bluegrass sod from heat in Jury. Crop Sci., 43:952-956.